

**Material**

10 staining jars, glass-rackets, kettle, tilting table

Prepare hot plate to 95-100°C

**Prepare** following solution serial in staining jars (200ml / cycle):

1. 0.2% SDS (Sigma L4522)
2. Aqua dest.
3. Prepare Aqua dest. in kettle for boiling
4. 70% Ethanol
5. 95% Ethanol
6. Isopropanol

Perform one cycle with 10 slides in a rack together

**Procedure**

1. Boil up water in kettle
2. Re-hydrate the spotted side of cDNA-arrays above the kettle and dry the slides for 10 sec on the hot plate
3. Incubate slides in 0.2% SDS for 2min while shaking (tilting table; change solution after 30 Slides)
4. Incubate slides in Aqua dest for 2min while shaking (tilting table; change solution after 10 Slides)
5. Incubate slides in boiling Aqua dest for 2min (change solution after 10 Slides)
6. Incubate slides in Ethanol serial, 70%igen Et-OH ( 10s ), 95%igen Et-OH (10s ) and Isopropanol (change solution after 30 Slides)
7. Dry slides continuously with nitrogen gun
8. Storage of the slides dry and dust-free in a box including slide number, spotting date and post-treatment date

**Quality control by test scanning**

Perform Prescan (PMT voltage settings: 532: 540; 635: 700; 40µm resolution) of every slide for quality control (background) and store information in folder named with post-treatment and spotting date; document background information in standard protocol sheet

